Özgün Araştırma

# Molecular Characterization of Mitochondrial Cytochrome c oxidase subunit 1 (Cox1) gene from Trichostrongylus Species (Nematoda: Trichostrongylidae) in Northern Iran

Kuzey İran'daki Trichostrongylus Türünden (Nematoda: Trichostrongylidae) Mitokondriyal Cytochrome c oxidase subunit 1 (Cox1) Geninin Moleküler Karakterizasyonu

🖸 Meysam Sharifdini<sup>1</sup>, 🔀 Elham Hajialilo<sup>2,3</sup>, 🕲 Hedayat Hosseinnezhad<sup>1</sup>, 🕲 Mohammad Ali Mohammadi<sup>4</sup>

<sup>1</sup>Guilan University of Medical Sciences, Department of Medical Parasitology and Mycology, Rasht, Iran
<sup>2</sup>Qazvin University of Medical Sciences, Medical Microbiology Research Center, Qazvin, Iran
<sup>3</sup>Qazvin University of Medical Sciences, Department of Parasitology and Mycology, Qazvin, Iran
<sup>4</sup>Kerman University of Medical Sciences, Research Center for Hydatid Disease in Iran, Kerman, Iran

Cite this article as: Sharifdini M, Hajialilo E, Hosseinnezhad H, Mohammadi MA. Molecular Characterization of Mitochondrial Cytochrome *c* oxidase subunit 1 (*Cox1*) gene from *Trichostrongylus* Species (Nematoda: Trichostrongylidae) in Northern Iran. Turkiye Parazitol Derg 2023;47(1):28-33.

# ABSTRACT

**Objective:** The objective of the present study was to identify *Trichostrongylus* species by molecular analysis and also phylogenetic relationships of *Trichostrongylus* species by mitochondrial Cytochrome c oxidase subunit 1 (*Cox1*) gene in Guilan province, northern Iran.

**Methods:** Abomasum and duodenum contents of 144 livestock were collected from sheep, goats, and cattle in Guilan province. Morphological survey was performed for initial screening. Total DNA was extracted, and the partial region of *Cox1* gene was amplified and sequenced. Genetic diversity was calculated and phylogenetic analysis of the data on nucleotide sequence was conducted by MEGA7 software.

**Results:** Three species of *Trichostrongylus* including *T. colubriformis, T. vitrinus, and T. axei* were identified by morphological characteristics. The genetic divergence within the species in the present study was observed for *T. axei* (0-2.5%), *T. colubriformis* (0.77%), and *T. vitrinus* (0%). The mean inter-species difference between the three species of *Trichostrongylus* obtained in this study was 14.4-15.4%.

**Conclusion:** The *Cox1* sequences of the members of *Trichostrongylus* spp. were highly variable and this could be used as a valuable measure to achieve a proper assessment on biodiversity. Sequence data generation from other species of *Trichostrongylus* will be needed to reconstruct the phylogenetic relationships of this genus of newtodes.

Keywords: Trichostrongylus spp., ruminants, Cox1, phylogenetic analysis, Iran

# ÖΖ

**Amaç:** Bu çalışmanın amacı, İran'ın kuzeyindeki Guilan eyaletindeki *Trichostrongylus* türlerinin moleküler analiz ve ayrıca mitokondriyal Cytochrome c oxidase subunit 1 (*Cox1*) geni ile *Trichostrongylus* türlerinin filogenetik ilişkilerinin belirlenmesidir. **Yöntemler:** Guilan ilindeki koyun, keçi ve sığırlardan 144 hayvanın abomasum ve duodenum içerikleri toplanmıştır. İlk tarama için morfolojik inceleme yapıldı. Toplam DNA ekstrakte edildi ve *Cox1* geninin kısmi bölgesi amplifiye edildi ve sekanslandı. Genetik çeşitlilik hesaplandı ve nükleotid dizisine ilişkin verilerin filogenetik analizi MEGA7 yazılımı ile yapıldı.

**Bulgular:** *T. colubriformis, T. vitrinus* ve *T. axei* dahil olmak üzere üç *Trichostrongylus* türü morfolojik özelliklerle tanımlanmıştır. Bu çalışmada türler içindeki genetik farklılık *T. axei* (%0-2,5), *T. colubriformis* (%0,77) ve *T. vitrinus* (%0) için gözlenmiştir. Bu çalışmada elde edilen üç *Trichostrongylus* türü arasındaki ortalama türler arası fark %14,4-15,4 olmuştur.



Received/Geliş Tarihi: 28.02.2022 Accepted/Kabul Tarihi: 19.09.2022

Address for Correspondence/Yazar Adresi: Elham Hajialilo, Qazvin University of Medical Sciences, Department of Parasitology and Mycology, Qazvin, Iran

Phone/Tel: +989127860343 E-mail/E-Posta: e.hajialilo@gmail.com ORCID ID: orcid.org/0000-0003-2159-4066

©Copyright 2023 Turkish Society for Parasitology - Available online at www.turkiyeparazitolderg.org ©Telif hakkı 2023 Türkiye Parazitoloji Derneği - Makale metnine www.turkiyeparazitolderg.org web sayfasından ulaşılabilir. **Sonuç:** *Trichostrongylus* spp. üyelerinin *Cox1* dizileri. oldukça değişkendi ve bu, biyoçeşitlilik hakkında uygun bir değerlendirme elde etmek için değerli bir ölçü olarak kullanılabilirdi. Bu nematod cinsinin filogenetik ilişkilerini yeniden yapılandırmak için diğer *Trichostrongylus* türlerinden dizi verisi oluşturulması gerekecektir.

Anahtar Kelimeler: Trichostrongylus spp., ruminantlar, Cox1, filogenetik analiz, İran

# **INTRODUCTION**

*Trichostrongylus* nematodes are highly prevalent and considered as gastrointestinal parasitic pathogens among ruminants with worldwide distribution (1,2). Clinical symptoms of humans are mild although in some patients gastrointestinal signs and eosinophilia may occur (3,4). These nematodes are major health challenges, causing reduced animal products (5,6). Several species of the parasite have been reported from herbivores with twelve valid species of species identified in humans (2,7,8).

Also, the frequency of Trichostrongylus spp. in human and animal hosts has been repeatedly reported in Iran (9-12). Ruminant infection was reported from various parts of Iran, with human infections found in Khuzestan, Isfahan, Tehran, Hormozgan, Kermanshah, Mazandaran, Guilan, Sistan & Baluchestan, and West Azerbaijan provinces (2,13-16). According to the morphological features reported in previous studies from Iran, several species of nematodes have been identified in human including T. orientalis, T. vitrinus, T. axei, T. colubriformis, T. probolurus, T. skrjabini, T. capricola, and T. lerouxi (9,10,15). In recent years, some studies clarified the human infections with T. vitrinus, T. axei, T. colubriformis, and T. longispicularis species by polymerase chain reaction (PCR) amplification of ITS2-rDNA region in endemic areas of northern Iran with T. colubriformis considered as the predominant species (2,8,13,17). Infection with various species of Trichostrongylus including T. colubriformis (11,12,15,18,19), T. vitrinus (11,12,15,18,19), T. axei (15), T. capricola (11,15), T. probolurus (11,12,15,18,19), T. longispicularis (11), T. orientalis (15), T. lerouxi (20), T. skrjabini (15), and T. hamatus (18) were reported in different herbivores such as sheep (11,12,15), goats (11,15), cattle (11,15), buffalos (11,15), and camels (18,19) in most parts of Iran. The predominant species of Trichostrongylus among different herbivores are T. colubriformis, T. vitrinus, and T. axei found in most parts of the country (15).

There is a tremendous diversity of the nematodes in the country (11,21) however, the molecular approaches, currently available and easily applicable, could accurately identify these species. Molecular studies based on ITS and 28S regions of ribosomal DNA were applied for genetic variation and phylogenetic analysis of super family Trichostrongyloidea (8,22-25). Although, numerous number of studies have focused on ITS2 for analysis of the Trichostrongylidae family in genetic variation, species detection, and phylogenetic relationships (2,8,21), yet mitochondrial (mt) genomes have the potential to present valuable information. Mt genomes are conserved and present large amounts of sequence data in the organisms, therefore mtDNA are used for evolutionary analyses, taxonomy, population genetics, and systematics studies (26-28). There are few studies that have investigated the mitochondrial gene of the Trichostrongylidae family, in which the mtDNA of Marshallagia marshalli, Haemonchus placei, Haemonchus contortus, T. vitrinus, T. axei, Ostertagia trifucata, and Teladorsagia circumcincta species were evaluated for phylogenetic relationship and species identification (26,29-34). The study explained to mitochondrial DNA diversity of O. ostertagi are five to ten times greater than typical estimates reported for species in other taxa (35). The population genetic structure and diversity of *H. contortus* by ribosomal and mitochondrial gene in Bangladesh showed low genetic differentiation but high gene flow among different populations of the parasite (36). Taxonomy studies of the nematodes based on sequences of coding mitochondrial genes are more accurate than non-coding ribosomal genes. While mitochondrial genomes are considered as suitable markers for population evolution studies (32,33), the studies targeting the mtDNA for identification of Trichostrongylidae family are very limited worldwide with even no single report on mitochondrial gene of the nematodes from Iran. Therefore, the present study focused on molecular phylogenetic analysis based on *Cox1* gene of *Trichostrongylus* species in northern Iran.

# **METHODS**

## **Sample Collection and Morphological Identification**

In current study, a total of 144 abomasum and duodenum specimens from livestock, including 72 cattle, 59 sheep, and 13 goats were collected from the abattoir of Talesh district in Guilan province, northern Iran during July to September 2018 (37).

The Trichostrongylidae family members were isolated by washing the abomasum and duodenum contents followed by passing through the 20, 40, and 100 mesh screens. The helminthes captured on mesh screens were examined under stereomicroscope. Morphological features were evaluated after cleaning the worms with normal saline and lactophenol. The samples were preserved in 70% ethanol at room temperature until used (38).

## **DNA Extraction and PCR Amplification**

Male parasites were isolated for DNA extraction. Selecting of male worms for molecular analysis was performed randomly. Total genomic DNA was extracted from one male worm of each species of trichostrongyloid nematodes collected from all study animals, using a commercial DNA extraction kit (Yekta Tajhiz Azma, Tehran, Iran) according to the manufacturer's instructions. The partial region of the Cox1 gene with approximately 700 bp was amplified using the LCO1490 HCO2198 (5'-GGTCAACAAATCATAAAGATATTGG-3') and (5'-TAAACTTCAGGGTGACCAAAAAATCA-3') sequences forward and reverse primers (39). The thermal PCR profiles included an initial denaturation step at 95 °C for 6 minutes followed by 35 cycles of denaturation at 95 °C for 45 seconds, annealing at 50 °C for 45 seconds, an initial extension step at 72 °C for 60 seconds, and a final extension step at 72 °C for 10 minutes.

## **Sequencing and Phylogenetic Analysis**

The PCR products were sequenced using an ABI 3130xl platform (Applied Biosystems, Foster City, California, USA). The sequences identified by the ABI system were edited and analyzed by BioEdit software (http://www.mbio.ncsu.edu/bioedit/bioedit.html).

The sequences were compared with the sequences deposited in the GenBank database by BLAST program (http://www.ncbi.nlm.nih. gov/). The sequences of the three species of genus *Trichostrongylus* derived from the domestic ruminants and deposited in the GenBank database were marked by the following Accession Numbers: MW051252-MW051254 for *T. axei*; MW051250 and MW051251 for *T. colubriformis*, and MW051255 and MW051256 for *T. vitrinus*.

Multiple sequence alignments were conducted by ClustalW incorporated in the BioEdit software. Phylogenetic tree was constructed by the MEGA7 software (Molecular and Evolution Genetic Analysis v7). The maximum likelihood method and the Tamura 3-parameter model were used for phylogenetic tree reconstruction. Bootstrap value was done based on 1000 replications in the topology of the tree. Genetic analysis of haplotypes was performed for Haplotype diversity, number of variable sites and number of haplotypes using DNAsp software (40).

# RESULTS

All of the study male worms were identified based on the morphological characteristics of male copulatory spicules and gubernaculum (Figure 1). *Trichostrongylus axei* was isolated from the cattle, sheep, and goats while *T. colubriformis* and *T. vitrinus* 

**Figure 1.** Posterior ends of males of *T. axei* (A), *T. vitrines* (B) and *T. colubriformis* (C)

were only detected among the sheep and goats. The isolates were successfully amplified for *Cox1* gene with specific band. The sequence results confirmed three species of *T. colubriformis*, *T. vitrinus*, and *T. axei* among the specimens. A dendrogram, based on the phylogenetic analysis, showed that the species were placed along, with the same species obtained from the GenBank database, into a distinct cluster of the tree (Figure 2). The genetic divergence within the species of *T. axei*, *T. colubriformis*, and *T. vitrinus* obtained in this study were 0-2.5%, 0.77%, and 0%, respectively. Two species of *Trichostrongylus* including *T. axei* and *T. vitrinus* isolated from the sheep and goats were quite similar. The intra-species distance rate within the specimens of *T. axei*, *T. colubriformis*, and *T. vitrinus* found in the present study and those available in the GenBank database amounted to 0.95-3.1% (1.9%), 0.19-4.08% (2.4%), and 0-2.32% (1.5%), respectively.

In this study, the mean inter-species differences between our *T. axei* specimens, compared with *T. colubriformis* and *T. vitrinus* isolates, were 14.4% and 14.6%, respectively. Also, the mean genetic difference between the *T. colubriformis* specimens was 15.4% when compared with *T. vitrinus*.

Based on our sequences and those deposited in the GenBank, the mean inter-species distance rates between the isolates of *T. axei* and those of *T. colubriformis* and *T. vitrinus* were 13.5% and 14.5%, respectively. Also, the mean genetic diversity between the isolates of *T. colubriformis* and those of *T. vitrinus* was 14.9%. The isolates were categorized into 5 haplotypes. The haplotype diversity for 7 isolates was calculated as Hd =0.9048. The distribution of *Trichostrongylus* isolates in each haplotypes is shown in Table 1.

# DISCUSSION

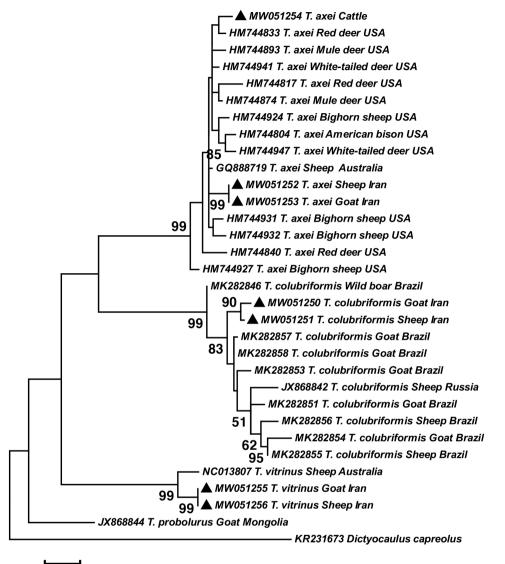
The three species of the *Trichostrongylus* including *T. colubriformis*, *T. vitrinus*, and *T. axei* identified in the present study, along with the data already reported from Iran confirm that the predominant species in herbivorous animals (15). *T. axei* was isolated from the cattle, sheep, and goats while *T. colubriformis* and *T. vitrinus* were only detected among the sheep and goats. *T. axei* (2.86%) was the most predominant nematodes of cattle in Ethiopia (41).

Iran is one of the most important foci for *Trichostrongylus* infection among human and animal hosts (12,16,42,43). Proper conditions such as humidity and climate in the northern parts of the country including Mazandaran and Guilan provinces lead to permanent establishment of the life cycle process of soil transmitted helminthes in the regions (2,8,42).

In the present study the authors used the sequence analysis protocol for detecting the mitochondrial *Cox1* gene, whereas several other studies, reported from Iran, employed *ITS-rDNA* gene specific for the phylogenetic analysis of *Trichostrongylus* species (2,8,13). The nuclear ribosomal gene is widely applied to the studies of deep and shallow phylogenetic relationship in the phylum Nematoda (2,8,44,45). Recent studies illustrated that the mitochondrial genes to be the proper options for phylogenetic approach and specifically for the *Cox1* gene that has mainly been used in population genetic surveys for various nematode parasites of the vertebrates (45-47).

Several studies have demonstrated that the sequence differences between the members of the *Trichostrongylus* are not noticeable when the detection protocol is based on the *ITS2* gene (2,8,13). Ashrafi et al. (13) reported a mean inter-species distance rate of 2.6% within different species of *Trichostrongylus* while in the current study the mean inter-species variation within our specimens and those available in the GenBank was 13.5-14.9%. Due to the high-level divergence in the *Cox1* gene, it could be considered as a valuable genetic tool for phylogenetic and taxonomic studies on the members of the *Trichostrongylus* genus. The phylogenetic tree constructed in our study represented that

the three species of *T. colubriformis*, *T. vitrinus*, and *T. axei* were separated in distinct cluster along with the same species obtained from other studies in different countries (Figure 2). The results of genetic diversity within the species showed that the intra-species distance rate among the present isolates was so close, indicating high proximity of the sequences in the region.



0.02

**Figure 2.** Phylogenetic tree of isolates of *Trichostrongylus* spp. obtained in this study (▲) and other isolates of *Trichostrongylus* spp. retrieved from GenBank based on *Cox1* gene. The tree was designed by using the maximum-likelihood test and the Tamura 3-parameter model as implemented in the MEGA7 software. *Dictyocaulus capreolus* was used as an out group

**Table 1.** Haplotypes groupings for *Trichostrongylus* isolates of partial cytochrome c oxidase gene subunit 1 (*Cox1*) and the accession numbers of isolates in each group

numbers of isolates in each group			
Haplotype	Number of isolates	Species	Accessions
Hap-1	1	T. colubriformis	MW051250
Hap-2	1	T. colubriformis	MW051251
Hap-3	2	T. axei	MW051252, MW051253
Hap-4	1	T. axei	MW051254
Hap-5	2	T. vitrinus	MW051255, MW051256

Little information on mitochondrial genes of Trichostrongyloidea superfamily is available. Palevich et al. (33) in New Zealand investigated the complete mitochondrial genomes of H. contortus and T. circumcincta by phylogenetic analysis. Another study, reported from Uzbekistan, was based on ribosomal (ITS2) and mitochondrial (Cox1) of Marshallagia sp. and concluded that the ITS2 sequences has little variation and is not a suitable gene for diagnosing different species, while Cox1 gene shows more diversities (32). Ostertagia trifurcata and Marshallagia marshalli were evaluated by phylogenetic analysis of the complete mitochondrial genes in China and the findings introduced complete mt genome sequence of the nematodes as a novel genetic marker for population genetic and molecular epidemiology (29,34). Two other studies, reported from Brazil and Australia, evaluated the complete mitochondrial genes of H. placei, T. circumcincta, T. vitrinus, and T. axei and suggested that the phylogenomics approach of mtDNA could be applied as a new genetic marker in phylogenetic analysis and geographic relationships among different isolates in population genetic studies (26,31). Moreover, the Cox1 and nad4 genes of T. axei were also analyzed for population genetic structure of the nematode in USA (30). Our study could be the basis for further sequence studies with greater sample sizes, especially the analysis of both nuclear and mitochondrial genes, are needed to provide a comprehensive understanding of the genetic variations of Trichostrongylus spp. in endemic areas and other parts of Iran.

# **CONCLUSION**

In the present study three species of *T. colubriformis, T. vitrinus,* and *T. axei* were observed among the specimens of Guilan province, northern Iran. This study concluded the genetic diversity of the *Cox1* gene is notable and the gene is suitable for analyzing the gene diversity of intra-species distance among helminthes. The scarcity of molecular data on *Cox1* gene within *Trichostrongylus* spp. in various geographical regions and hosts makes it necessary to produce sufficient data on diversities of this gene which eventually leads to reconstruct the total phylogenetic relationships of this group of nematode. Thus, the findings of the present study suggest that the analysis of complete mitochondrial genome to be the focus of further experiments in the future research.

#### \* Acknowledgements

We would like to appreciate the assistance offered by the colleagues at the Department of Parasitology and Mycology, Medical School, Guilan University of Medical Sciences. We thank Dr. Ali-Asghar Pahlevan for editing the final version of the English manuscript. The authors are grateful to the abattoir personnel in Talesh, Guilan Province of Iran.

## \* Ethics

**Ethics Committee Approval:** This study was approved by the Medical Ethics Committee of Guilan University of Medical Sciences (Approval code: IR.GUMS.REC.1397.176).

## Informed Consent: Not applicable.

Peer-review: Internally and externally peer-reviewed.

## \* Authorship Contributions

Concept: M.S., Design: M.S., E.H., Data Collection or Processing: H.H., Analysis or Interpretation: M.S., M.A.M., Literature Search: E.H., Writing: E.H., Critical Review: M.S., E.H.

**Conflict of Interest:** No conflict of interest was declared by the authors

**Financial Disclosure:** This study was financially supported by Guilan University of Medical Sciences.

#### REFERENCES

- 1. Roberts L, Janovy J. Foundations of parasitology. 8th edn. 701 pp. New York, McGraw-Hill; 2009.
- Sharifdini M, Heidari Z, Hesari Z, Vatandoost S, Kia EB. Molecular phylogenetics of *Trichostrongylus* species (Nematoda: Trichostrongylidae) from humans of Mazandaran province, Iran. Korean J Parasitol 2017; 55: 279-85.
- Ghanbarzadeh L, Saraei M, Kia E, Amini F, Sharifdini M. Clinical and haematological characteristics of human trichostrongyliasis. J Helminthol 2018; 93: 149-53.
- Wall EC, Bhatnagar N, Watson J, Doherty T. An unusual case of hypereosinophilia and abdominal pain: an outbreak of Trichostrongylus imported from New Zealand. J Travel Med 2011; 18: 59-60.
- da Rocha LO, da Silva Lemos GC, Vieira IJC, Braz-Filho R, de Paiva Freitas S, Glória LS, et al. Chemical characterization and *in vitro* biological activity of *Cymbopogon citratus* extracts against *Haemonchus* spp. and Trichostrongylus spp. nematodes from sheep. Parasitology 2020; 147: 1559-68.
- McLeod R. Costs of major parasites to the Australian livestock industries. Int J Parasitol 1995; 25: 1363-7.
- Phosuk I, Intapan PM, Sanpool O, Janwan P, Thanchomnang T, Sawanyawisuth K, et al. Molecular evidence of Trichostrongylus colubriformis and Trichostrongylus axei infections in humans from Thailand and Lao PDR. Am J Trop Med Hyg 2013; 89: 376-9.
- Sharifdini M, Derakhshani S, Alizadeh SA, Ghanbarzadeh L, Mirjalali H, Mobedi I, et al. Molecular identification and phylogenetic analysis of human Trichostrongylus species from an endemic area of Iran. Acta Trop 2017; 176: 293-9.
- Ghadirian E. Human infection with Trichostrongylus lerouxi (Biocca, Chabaud, and Ghadirian, 1974) in Iran. Am J Trop Med Hyg 1977; 26: 1212-3.
- Ghadirian E, Arfaa F, Sadighian A. Human infection with Trichostrongylus capricola in Iran. Am J Trop Med Hyg 1974; 23: 1002-3.
- Ghasemikhah R, Mirhendi H, Kia E, Mowlavi G, Sarmadian H, Meshgi B, et al. Morphological and morphometrical description of Trichostrongylus species isolated from domestic ruminants in Khuzestan province, southwest Iran. Iran J Parasitol 2011; 6: 82.
- Shahbazi A, Fallah E, Koshki MK, Nematollahi A, Ghazanchaei A, Asfaram S. Morphological characterization of the Trichostrongylus species isolated from sheep in Tabriz, Iran. Research Opinions in Animal and Veterinary Sciences 2012; 2: 309-12.
- Ashrafi K, Sharifdini M, Heidari Z, Rahmati B, Kia EB. Zoonotic transmission of Teladorsagia circumcincta and Trichostrongylus species in Guilan province, northern Iran: molecular and morphological characterizations. BMC Infect Dis 2020; 20: 1-9.
- Ashrafi K, Tahbaz A, Sharifdini M, Mas-Coma S. Familial Trichostrongylus infection misdiagnosed as acute fascioliasis. Emerg Infect Dis 2015; 21: 1869.
- Ghadirian E, Arfaa F. Present status of trichostrongyliasis in Iran. Am J Trop Med Hyg 1975; 24: 935-41.
- 16. Sharifdini M, Ghanbarzadeh L, Barikani A, Saraei M. Prevalence of intestinal parasites among rural inhabitants of Fouman, Guilan Province,

Northern Iran with emphasis on Strongyloides stercoralis. Iran J Parasitol 2020; 15: 91.

- 17. Gholami S, Babamahmoodi F, Abedian R, Sharif M, Shahbazi A, Pagheh A, et al. Trichostrongylus colubriformis: possible most common cause of human infection in Mazandaran province, North of Iran. Iran J Parasitol 2015; 10: 110-5.
- Anvari-Tafti M, Sazmand A, Hekmatimoghaddam S, Moobedi I. Gastrointestinal helminths of camels (Camelus dromedarius) in center of Iran. Trop Biomed 2013; 30: 56-61.
- Borji H, Razmi GR, Movasaghi A, Naghibi AA, Maleki M. S A study on gastrointestinal helminths of camels in Mashhad abattoir, Iran. Iranian Journal of Veterinary Research, Shiraz University 2010; 11: 174-9.
- Biocca E, Chabaud A, Ghadirian E. [Trichostrongylus lerouxi n. sp., parasite of Bos taurus]. Parassitologia 1974; 16: 199-207.
- 21. Ghasemikhah R, Sharbatkhori M, Mobedi I, Kia E, Harandi MF, Mirhendi H. Sequence analysis of the second internal transcribed spacer (ITS2) region of rDNA for species identification of Trichostrongylus nematodes isolated from domestic livestock in Iran. Iran J Parasitol 2012; 7: 40-6.
- de Bellocq JG, Ferte H, Depaquit J, Justine JL, Tillier A, Durette-Desset MC. Phylogeny of the Trichostrongylina (Nematoda) inferred from 28S rDNA sequences. Mol Phylogenet Evol 2001; 19: 430-42.
- 23. Hoberg EP, Monsen KJ, Kutz S, Blouin MS. Structure, biodiversity, and historical biogeography of nematode faunas in holarctic ruminants: morphological and molecular diagnoses for Teladorsagia boreoarcticus n. sp.(Nematoda: Ostertagiinae), a dimorphic cryptic species in muskoxen (Ovibos moschatus). J Parasitol 1999; 85: 910-34.
- von Samson-Himmelstjerna G, Harder A, Schnieder T. Quantitative analysis of ITS2 sequences in trichostrongyle parasites. Int J Parasitol 2002; 32: 1529-35.
- Pandi M, Sharifdini M, Ashrafi K, Atrkar Roushan Z, Rahmati B, Hajipour N. Comparison of Molecular and Parasitological Methods for Diagnosis of Human Trichostrongylosis. Front Cell Infect Microbiol 2021; 11: 759396.
- 26. dos Santos LL, Prosdocimi F, Lima NCB, da Costa IR, Cardoso DC, Drummond MG, et al. Comparative genomics and phylogenomics of Trichostrongyloidea mitochondria reveal insights for molecular diagnosis and evolutionary biology of nematode worms. Gene Reports 2017; 9: 65-73.
- Hu M, Chilton NB, Gasser RB. The mitochondrial genomics of parasitic nematodes of socio-economic importance: recent progress, and implications for population genetics and systematics. Adv Parasitol 2004; 56: 134-213.
- Saccone C, De Giorgi C, Gissi C, Pesole G, Reyes A. Evolutionary genomics in Metazoa: the mitochondrial DNA as a model system. Gene 1999; 238: 195-209.
- 29. Ahmad AA, Yang X, Zhang T, Wang C, Zhou C, Yan X, et al. Characterization of the complete mitochondrial genome of *Ostertagia trifurcata* of small ruminants and its phylogenetic associations for the Trichostrongyloidea superfamily. Genes (Basel) 2019; 10: 107.
- Archie EA, Ezenwa VO. Population genetic structure and history of a generalist parasite infecting multiple sympatric host species. Int J Parasitol 2011; 41: 89-98.
- Jex AR, Hall RS, Littlewood DTJ, Gasser RB. An integrated pipeline for next-generation sequencing and annotation of mitochondrial genomes. Nucleic Acids Res 2010; 38: 522-33.
- 32. Kuchboev A, Sobirova K, Karimova R, Amirov O, von Samson-Himmelstjerna G, Krücken J. Molecular analysis of polymorphic species

of the genus Marshallagia (Nematoda: Ostertagiinae). Parasit Vectors 2020; 13: 411.

- Palevich N, Maclean PH, Choi YJ, Mitreva M. Characterization of the complete mitochondrial genomes of two sibling species of parasitic roundworms, *Haemonchus contortus* and *Teladorsagia circumcincta*. Front Genet 2020; 11: 573395.
- 34. Sun MM, Han L, Zhang FK, Zhou DH, Wang SQ, Ma J, et al. Characterization of the complete mitochondrial genome of Marshallagia marshalli and phylogenetic implications for the superfamily Trichostrongyloidea. Parasitol Res 2018; 117: 307-13.
- Dame JB, Blouin MS, Courtney CH. Genetic structure of populations of Ostertagia ostertagi. Vet Parasitol 1993; 46: 55-62.
- Dey AR, Zhang Z, Begum N, Alim A, Hu M, Alam MZ. Genetic diversity patterns of Haemonchus contortus isolated from sheep and goats in Bangladesh. Infect Genet Evol 2019; 68: 177-84.
- Hosseinnezhad H, Sharifdini M, Ashrafi K, Atrkar Roushan Z, Mirjalali H, Rahmati B. Trichostrongyloid nematodes in ruminants of northern Iran: prevalence and molecular analysis. BMC Vet Res 2021; 17: 371.
- Barghandan T, Hajialilo E, Sharifdini M, Javadi A. Prevalence and phylogenetic analysis of gastrointestinal helminths (Nematoda: Trichostrongylidae) in ruminant livestock of northwest Iran. Ankara Üniv Vet Fak Derg 2019; 67: 65-72.
- Folmer O, Black M, Hoeh W, Lutz R, Vrijenhoek R. DNA primers for amplification of mitochondrial cytochrome c oxidase subunit I from diverse metazoan invertebrates. Mol Mar Biol Biotechnol 1994; 3: 294-9.
- Librado P, Rozas J. DnaSP v5: a software for comprehensive analysis of DNA polymorphism data. Bioinformatics 2009; 25: 1451-2.
- Abuhay M, Hamid M, Tintagu T. Prevalence and Species Composition of Abomasal Nematodes of Cattle slaughtered at Abergelle Export Abattoir, Mekelle, Ethiopia. International Journal of Veterinary Science & Technology 2018; 2: 33-7.
- Alemi A, Arfaa F. Prevalence of intestinal helminthiasis in the rural area of gilan province (caspian littoral). Iranian Journal of Public Health 1978; 25-34.
- Massoud J, Arfaa F, Jalali H, Keyvan S. Prevalence of intestinal helminths in Khuzestan, Southwest Iran, 1977. Am J Trop Med Hyg 1980; 29: 389-92.
- 44. Holterman M, van der Wurff A, van den Elsen S, van Megen H, Bongers T, Holovachov O, et al. Phylum-wide analysis of SSU rDNA reveals deep phylogenetic relationships among nematodes and accelerated evolution toward crown clades. Mol Biol Evol 2006; 23: 1792-800.
- 45. Kiewnick S, Holterman M, van den Elsen S, van Megen H, Frey JE, Helder J. Comparison of two short DNA barcoding loci (COI and COII) and two longer ribosomal DNA genes (SSU & LSU rRNA) for specimen identification among quarantine root-knot nematodes (*Meloidogyne* spp.) and their close relatives. European Journal of Plant Pathology 2014; 140: 97-110.
- Nadler SA, Hudspeth DS. Phylogeny of the Ascaridoidea (Nematoda: Ascaridida) based on three genes and morphology: hypotheses of structural and sequence evolution. J Parasitol 2000; 86: 380-93.
- 47. Otranto D, Testini G, De Luca F, Hu M, Shamsi S, Gasser R. Analysis of genetic variability within Thelazia callipaeda (Nematoda: Thelazioidea) from Europe and Asia by sequencing and mutation scanning of the mitochondrial cytochrome c oxidase subunit 1 gene. Mol Cell Probes 2005; 19: 306-13.